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**Trace Metals in Airborne Particulate Matter and Genomic Characterization of Associated Microorganisms: Insights into Health Effects from an Industrialized, Near-Roadway Site in Houston**

**Abstract**

This research simultaneously measured major/trace metals and microorganism diversity in airborne particulate matter ≤ 10 µm in aerodynamic diameter. The objectives were to (i) analyze the elemental composition of PM$_{10}$, (ii) perform source apportionment to quantify vehicular contributions, and (iii) implement state-of-the-art next generation sequencing tools to evaluate airborne microorganism diversity and prevalence. Filter samples were collected over a 9-day period spanning August 10–August 18, 2018, at Clinton Drive in Houston, Texas (latitude 29.73372; longitude −95.25759). Source apportionment modeling resolved vehicular emissions, resuspended local soil/road dust, and construction activities as major PM$_{10}$ sources. Coincidentally, our sampling campaign captured a strong African dust event in Houston. Hence, all our results include a foreign component of aerosol mass, chemistry, and microbiology. Estimated vehicular emissions ranged between ~4.6–11.2 µg/m$^3$, averaging 7 µg/m$^3$. This constituted ~11 percent of the measured total PM$_{10}$ mass on average. Estimated contributions from local soil and road dust were between ~8.0–28.5 µg/m$^3$, averaging 19.9 µg/m$^3$. This constituted ~31 percent of the measured total PM$_{10}$ mass on average. Opportunistic human, plant, or animal pathogenic bacterial species were identified including *Escherichia coli*, *Propionibacterium acnes*, *Roseomonas mucosa*, and *Haemophilus parainfluenzae*. Several genera listed in the World Health Organization’s (WHO’s) global priority pathogens list of multidrug and antibiotic-resistant bacteria and tuberculosis were detected including *Acinetobacter*, *Enterococcus*, *Haemophilus*, *Mycobacterium*, *Pseudomonas*, and *Staphylococcus*. Fungi responsible for invasive human diseases such as *Aspergillus fumigatus*, *Fusarium* spp., and *Talaromyces* spp. that are listed as being priority pathogens for global public health by WHO appeared in nearly every sample along with all four most prominent allergenic fungal genera, viz. *Alternaria*, *Cladosporium*, *Aspergillus*, and *Penicillium*.

**Key Words**

PM$_{10}$, Elemental analysis, Airborne bacteria, Airborne Fungi, Bioaerosols

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Executive Summary

Problem statement
Traffic related air pollution (TRAP) diminishes lung function, induces asthma, allergic rhinitis, respiratory infections, retards brain function, and stunts children’s overall development [1-5]. Additionally, a high incidence of lung cancers, allergies, pulmonary and cardiovascular diseases, and premature death has been reported in populations living adjacent to highways and busy roadways [4]. Since particulate matter [6] mass concentrations are strongly correlated to morbidity and mortality [7, 8], a portion of the negative health impacts attributed to TRAP is assigned to inhalable particulate matter (PM). It is emphasized that causal correlations are well-accepted only for PM mass (not its specific constituents), justifying federal PM10 and PM2.5 National Ambient Air Quality Standards based on daily or annual average mass concentrations (μg of PM per m3 of air).

An important data gap in the existing literature is the detailed characterization of long-term exposure to lower levels of a wide range of vehicular metals that potentially lead to chronic health issues. In this research, we addressed this lack of rigorous information traffic related PM components by measuring concentrations of ~50 elements in vehicular emissions using mass spectrometry and then performing source apportionment modeling. Our measurements and calculations incorporate realistic estimates of primary tailpipe and non-tailpipe PM10 (particulate matter with aerodynamic diameters ≤10 μm) emissions under actual on-road conditions, including several real-world sources of variability such as maintenance histories, vehicle age, engine size and type, and driving habits that were obtained from measurements near a heavily trafficked roadway [9, 10].

Technical objectives, approach, and methodology
Our overarching objectives of this first comprehensive study designed to simultaneously measure major/trace metals and microorganism diversity in airborne coarse particulate matter were to:

- Quantify vehicular contributions to PM10 and associated human exposure by analyzing aerosols’ elemental composition and perform source apportionment using the United States Environmental Protection Agency’s Chemical Mass Balance (CMB v8.2) model.
- Implement state-of-the-art next generation sequencing tools to evaluate airborne microorganism diversity and prevalence in PM10 samples.

Filter samples were collected over a 9-day period spanning August 10–August 18, 2018, at Clinton Drive in Houston, Texas (latitude 29.73372; longitude −95.25759). This site is in a densely populated and economically depressed area (26.4 percent poverty rate) inhabited largely by ethnic minorities (91 percent Hispanics, African Americans, American Indians, and Alaska Natives) bringing environmental justice issues into focus. Clinton Drive is close to several heavily trafficked roads including Interstate-610 and Interstate-10, which had annual average daily traffic counts of 178,800 and 194,945 vehicles, respectively, and is within the Houston Ship Channel region, a hyper-industrialized section of the city. Therefore, air quality indices are frequently in the unhealthy range at this site, which consequently has been the target of many investigations [11-14].

One half of each filter was analyzed for elemental concentrations using high temperature microwave assisted acid digestion and inductively coupled plasma–mass spectrometry. The other half was used to quantify microorganism diversity by amplifying the V3 and V4 hypervariable regions of 16S ribosomal DNA (rDNA) and the highly variable internal transcribed spacer (ITS) region of rDNA (18S/ITS). Bioinformatics tools were used to identify the composition and abundance of prokaryotes and eukaryotes using 16S and 18S/ITS data, respectively.

Key findings
Source apportionment modeling resolved vehicular emissions, resuspended local soil/road dust, and construction activities as major PM10 sources. Coincidentally, our sampling campaign captured a strong African dust event in
Houston. Hence, all our results include a foreign component of aerosol mass, chemistry, and microbiology. The main findings from chemical mass balance modeling were:

- Measured ambient PM$_{10}$ concentrations ranged between $\sim$45–116 µg/m$^3$, averaging 73 µg/m$^3$.
- Estimated vehicular emissions ranged between $\sim$4.6–11.2 µg/m$^3$, averaging 7 µg/m$^3$. This constituted $\sim$11 percent of the measured total PM$_{10}$ mass on average.
- Estimated contributions from local soil and road dust were between $\sim$8.0–28.5 µg/m$^3$, averaging 19.9 µg/m$^3$. This constituted $\sim$31 percent of the measured total PM$_{10}$ mass on average.
- Estimated contributions from construction activities were between $\sim$1.5–18.1 µg/m$^3$, averaging 6.3 µg/m$^3$. This constituted $\sim$31 percent of the measured total PM$_{10}$ mass on average.
- Estimated contributions from North African dust were between $\sim$2.0–65.7 µg/m$^3$, averaging 24.5 µg/m$^3$. This constituted $\sim$8.5 percent of the measured total PM$_{10}$ mass on average.

The main findings from genomic sequencing of ambient PM$_{10}$ were:

- Opportunistic human, plant, or animal pathogenic bacterial species were identified including *Escherichia coli*, *Propionibacterium acnes*, *Roseomonas mucosa*, and *Haemophilus parainfluenzae*. Several genera corresponding to human and plant pathogens were also detected.
- Several genera listed in the World Health Organization’s (WHO) global priority pathogens list of multidrug and antibiotic-resistant bacteria and tuberculosis [15] were detected including *Acinetobacter*, *Enterococcus*, *Haemophilus*, *Mycobacterium*, *Pseudomonas*, and *Staphylococcus*.
- Fungi responsible for invasive human diseases such as *Aspergillus fumigatus*, *Fusarium* spp. and *Talaromyces* spp. that are listed as being priority pathogens for global public health by WHO [16] appeared in nearly every sample along with all four most prominent allergenic fungal genera, viz. *Alternaria*, *Cladosporium*, *Aspergillus*, and *Penicillium*.
- Numerous extremophilic and thermophilic genera of bacteria (*Chroococcidiopsis*, *Deinococcus*, *Hydrogenophilus*, *Meiothermus*, and *Saccharomonospora*) and fungi (*Alternaria*, *Aspergillus*, *Chaetomium*, *Cladosporium*, *Coprinopsis*, *Pencillium*, *Rhizopus*, *Scytalidium*, *Stemphylium*, *Talaromyces*, *Thanatephorus*, *Thermomyces*, *Thielavia*, and *Wallemia*) were identified [17].

**Project Impacts**

This research is a systematic and rigorous investigation that coupled primary PM$_{10}$ sources, detailed elemental characterization, and DNA sequencing to provide essential information for animal, plant, and ecosystem health studies in a North American metroplex. We identified transportation sources, local industrial sources, and long-range transported dust from the Sahara-Sahel region impacting the chemical and microbiological composition of respirable aerosols in Houston, TX. We published our major findings in the journal *Environmental Science & Technology* [18]. Our outreach activities were geared towards informing elementary school students about environmental impacts of the transportation sector. Demonstrations performed for fourth graders familiarized them with direct (primary) emissions from motor vehicles, focusing their attention on environmental issues. This research addresses two of CARTEEH’s priority areas: measurement and modeling, in the context of public health. Results from this short-term pilot-study provide the foundation for designing future investigations to better demarcate long-range transported microorganisms from locally aerosolized ones. Future research also needs to complement metagenomically identified bacteria and fungi by monitoring viability to better assess microorganisms’ role in ecosystem and animal health.
Acknowledgments

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Background and Introduction

Populations living adjacent to highways and busy roadways have an increased risk for morbidity and premature mortality [19-22] with reports of a high incidence of lung cancers, allergies, respiratory and cardiovascular diseases, and even premature death [23]. A portion of these health impacts has been assigned to airborne particulate matter [6] and its metals content [8, 19]. This necessitates detailed elemental characterization of vehicular particulate matter (PM) emissions and accurate identification and apportionment of ambient PM arising from motor vehicles. For these and other impacts of aerosols on human and ecosystem health [24-32], much research has focused on characterizing their chemical composition and epidemiological/toxicological effects to develop scientific policy enabling targeted emission control strategies to reduce their mass concentrations [31, 33, 34].

In addition to direct emissions, motor vehicles also resuspend soil and road dust that harbor microorganisms, indirectly emitting toxins into the local atmosphere, which may induce negative health impacts. In this research, we targeted PM$_{10}$ to capture fungi and several other airborne microbes even though PM$_{2.5}$ and finer fractions penetrate deeper into the respiratory tract. One of the novelties of our research lies in the fact that in contrast to physical and chemical characterization of ambient aerosols, comparatively fewer reports have considered airborne microorganisms even though they also influence public health and the environment [17, 24, 35, 36].

For these reasons, we undertook a pilot study of aerosols enriched in metals and microorganisms. We focused on the city of Galena Park, which is adjacent to the Houston Ship Channel that serves the United States’ largest petrochemical complex, the second busiest port by cargo volume, and myriad other concatenating industries. Galena Park is predominantly populated by Hispanics, African Americans, and Native Americans (92 percent) with a low per capita income (only $15,190) and high (26.4 percent) poverty rate [37]. Since air pollution exposure is skewed to low income and minority populations [38], our work also addressed environmental justice concerns [39-41]. Further, Houston is an interesting urban testbed to evaluate traffic related aerosols since Texas ranks second only to California in terms of vehicle miles traveled. Hence, the explicit focus of the work was to measure a wide suite of particulate metals and microorganism diversity in airborne PM$_{10}$ at a busy intersection near the ship channel.

The overarching objectives of this first-of-its-kind study designed to simultaneously measure major/trace metals and microorganism diversity in airborne coarse particulate matter in North America were to:

- Collect ambient PM$_{10}$ samples and quantify elemental concentrations using high temperature microwave assisted acid digestion and inductively coupled plasma–mass spectrometry (ICP-MS),
- Perform source apportionment using the United States Environmental Protection Agency’s (EPA’s) Chemical Mass Balance (CMB v8.2) model to determine the extent to which various sources including tailpipe and non-tailpipe emissions and resuspended road dust contributed to ambient PM$_{10}$ concentrations,
- Implement state-of-the-art next generation sequencing tools to evaluate airborne microorganism diversity and prevalence in PM$_{10}$ samples, and
- Develop outreach activities geared towards informing elementary school students about environmental impacts of the transportation sector.

Problem

As explained above, it is well recognized that traffic related air pollution causes a wide range of human health problems. Even though PM [6] regulations are mass based, it is well-accepted that metals are an important component that is responsible for the observed health end points. Hence, it is important to quantitatively estimate
aerosols emitted by motor vehicles and evaluate their metals composition. An additional consideration is that although several studies have measured metals in airborne PM, comparatively fewer reports have considered airborne microorganisms even though they also influence public health and the environment.

**Approach**

We undertook a systematic and rigorous study to measure particulate metals and airborne microorganisms at a heavily trafficked intersection in greater Houston. Filter PM_{10} samples were collected over a 9-day period spanning August 10–August 18, 2018, at Clinton Drive in Houston, Texas (latitude 29.73372; longitude −95.25759). They were analyzed for metals using ICP-MS following high temperature microwave assisted acid digestion. This information was used to perform source apportionment to quantify PM emissions from motor vehicles, resuspended soil and road dust, and other local and global sources that influenced ambient aerosol concentrations. Bacterial and fungal diversity were quantified by amplifying the V3 and V4 hypervariable regions of 16S ribosomal DNA (rDNA) and the highly variable internal transcribed spacer (ITS) region of rDNA (18S/ITS).

**Methodology**

**PM_{10} sampling**

A ThermoFisher 2025i Partisol sequential air sampler was used to collect samples on PTFE filters. All sampler parts (i.e., inlets, filter cassettes, filter cartridge, carrier box support, and filters) were disinfected with 70 percent ethyl alcohol followed by 5 percent hypochlorite solution. Filters were disinfected by exposing them to UV light for 15 minutes inside a biosafety cabinet (Baker SterilGARD, Class II Type A2). Filters were refrigerated at −80°C immediately after removing from the sampler (using sterile tweezers), weighed to measure PM_{10} mass concentrations, cut into two halves using a sterilized ceramic knife inside a biosafety cabinet (one for elemental analysis and the other for sequencing), and again stored at −80 °C in sterile petri dishes until further analysis.

**Elemental analysis**

The filter half designated for elemental analysis was digested first in 3 mL of trace metal grade concentrated HNO₃ with additional optima grade concentrated HF in a ratio of 0.3 mL for every 10 mg of sample, based on our previous work [12, 13, 42, 43]. This was followed by adding 5 percent w/v boric acid solution to mask the insoluble fluoride complexes and was re-digested. Each stage was performed at 200°C and 200 psig in a microwave oven (MARS 6, CEM Corporation) for a dwell time of 20 min. The digested aliquot was then diluted to 2 percent nitric acid matrix before analyzing with ICP-MS. The 46 elements were measured including 31 from Groups 1–16 (Na, Mg, Al, Si, K, Ca, Sc, Ti, V, Cr, Mn, Fe, Co, Ni, Cu, Zn, Ga, As, Se, Rb, Sr, Zr, Mo, Cd, Sn, Sb, Cs, Ba, Pb, Th, and U) and 15 rare earths (Y, La, Ce, Pr, Nd, Sm, Eu, Gd, Tb, Dy, Ho, Er, Tm, Yb, and Lu). Al, V, Cr, Fe, Ni, Cu, and Zn were analyzed in Dynamic Reaction Cell mode using ammonia as the cell gas to reduce polyatomic interferences [12, 42-45].

**Nucleic acid extraction and gene sequencing**

Nucleic acids were extracted from the Teflon filters using bead-beating and purification with centrifugal filters. PM_{10} filters were cut in half, folded with sterile forceps, and placed into 2-mL bead beating tubes (National Scientific supply, BC20NA-PS), along with 200 mg of ≤ 106 μm diameter glass beads (Sigma, G-4649), 200 mg of 425–600 μm glass beads (Sigma, G-8772), and 650 μL of sodium phosphate buffer prepared aseptically by mixing 1.0 μL Tween 20 with 10 mL of 0.1 M sodium phosphate/10 mM EDTA (Teknova, Hollister, CA). The bead beating tube was mechanically disrupted with a mini-bead beater (BioSpec Products, Bartlesville, OK) for 3 min at 3450 oscillations/min and subsequently placed on ice for 5 min. After cooling, tubes were centrifuged at 7000 × g for 2 min. 500 μL of supernatant was transferred to an Ultrafree-MC centrifugal filter (0.22 μm pore size, hydrophilic PVDF, 0.5 mL volume, UFC30GV05) and centrifuged at 10,000 × g for 3 min. Following the centrifugal pre-filtration, the filtrate was transferred to an Amicon Ultra-0.5 (100,000 Molecular Weight Cut-Off) centrifugal filter device.
(YM-100, Millipore) and centrifuged for 3 min at 7000 × g. This step concentrated nucleic acids on the filter while allowing inhibitory compounds to pass through. The filter was first washed twice, each time by adding 200 μL of TE buffer to the retentate cup and centrifuging for 3 min at 7000 × g. After the second wash, 100 μL of the TE was added and centrifuged for 1 min at 7000 × g and subsequently in 10 second pulses as needed to recover 100 μL through the filter. Nucleic acids were recovered from the top of the Amicon filter insert by inverting the filter into a fresh tube by centrifugation at 1000 × g for 2 min. Final purification of nucleic acids was performed as per manufacturer’s protocol using a polyvinylpolypyrrolidone spin column (Spin-IV-HRC, Zymo Research).

We followed the protocol recommended by Swift Biosciences to generate amplicons that cover all variable regions of the 16S rDNA, ITS1, and ITS2 genes in a single primer pool using the Swift Amplicon 16S + ITS kit. The PCR products (obtained following their protocol) were cleaned up using the Agencourt AMPure XP beads (Beckman Coulter Genomics), and the purified amplicon was resuspended in 25 μl TE buffer. The first round of purified amplicon was amplified using the Illumina adaptors specific dual indexed Nextera XT barcoded primers of index 1 (i7) adapter and index 2 (i5) adapter with 15 cycles of amplification followed by clean-up. The amplicons were purified with Agencourt AMPure XP beads, quantified with Qubit (Invitrogen), and verified the amplicon size with Agilent TapeStation 2200 system. The library pool was diluted to obtain a final concentration of 8 pM. The 600 μL of the library pool was loaded onto a MiSeq v2 reagent cartridge (500 cycle v2 kit) and 251 bp paired-end sequencing protocol (2 × 251 cycles) was performed on MiSeq platform (Illumina).

Raw sequence reads were cleaned and processed using QIIME v1.9.1 [46]. Forward and reverse reads were merged, assigned to samples according to barcodes, and then trimmed to remove barcode and primer sequences. Sequences shorter than 200 bp, those containing ambiguous bases, and those with mean quality scores of < 20 were discarded. The remaining sequences were then aligned to the reference UCHIME RDP ‘Gold’ database to identify and remove chimera sequences. Sequence reads were clustered into Operational Taxonomic Units (OTUs) using VSEARCH v.1.9.6[47] and the SILVA 119 database[48] for 16s and the UNITE database for ITS[49]. For both gene regions, sequences were assigned to OTUs using a 0.8 threshold, with taxonomic categories parsed up to the species level, and subsequently rarified prior to downstream analyses.

Source apportionment
EPA’s Chemical Mass Balance (CMB v8.2) model was employed to quantify contributions of various sources to PM10 mass concentrations [12, 13, 50, 51]. Na, Mg, Si, K, Ca, Ti, Co, Cu, As, Se, Sr, Sn, Sb, Mo, Ba, Pb, Al, V, Cr, Fe, Ni, Zn, Y, La, Ce, Pr, Nd, and Sm were chosen as fitting species based on their importance as elemental tracers and level of uncertainty in our laboratory measurements. Pb, Cd, Sn, Pb, Mo, As, and Zn were selected as traffic-related metals [43, 51, 52]; Zn, Pb, Cr, Mn, Mg, Co, and Ni were selected to track high-temperature industrial processes [45]; rare earths (La, Ce, Nd, Pr, Sm, Gd, and Eu) were chosen for petroleum refining (fluidized bed catalytic cracking) activities and crustal matter [14, 43, 44]; V and Ni were chosen for oil combustion and shipping activities [53, 54]; and Al, Si, Ti, Ca, Fe and Y were used to isolate crustal mass [13, 43].

Results
Overall trends in PM10 and PM2.5 mass
Details of the nine daily PM10 samples (labelled S1–S9) collected are given below in Table 1. PM2.5 concentrations that were measured at the same location by the Texas Commission on Environmental Quality (TCEQ) are also included.

<table>
<thead>
<tr>
<th>Sample ID</th>
<th>Start date</th>
<th>End date</th>
<th>PM10 (µg/m³)</th>
<th>PM2.5 (µg/m³)</th>
<th>Apportioned vehicular emissions (µg/m³)</th>
<th>Apportioned local soil and road dust emissions (µg/m³)</th>
</tr>
</thead>
</table>
As summarized in Table 1, PM$_{2.5}$ and PM$_{10}$ exhibited similar trends increasing ~3-fold from the first sample (S1) to the third sample (S3) before progressively declining to routine levels in the last sample (S9). The PM$_{2.5}$ level was 37.9 µg/m$^3$ in sample S3, which is more than three times the annual average primary standard of 12 µg/m$^3$ established by EPA. The average PM$_{2.5}$ concentration throughout this sampling period was 19 µg/m$^3$, which is 58 percent higher than EPA’s annual average primary National Ambient Air Quality Standard (NAAQS). It even exceeded the primary and secondary 24-hour NAAQS standard of 35 µg/m$^3$. PM$_{10}$ never exceeded the 24-hour primary and secondary NAAQS but approached the limit of 150 µg/m$^3$ in sample S3. PM$_{10}$ averaged 72 µg/m$^3$ over the study period.

Trends in elemental concentrations

Figure 1 summarizes elemental concentrations in PM$_{10}$ over the study duration. As seen, elemental concentrations varied over nearly seven orders of magnitude (e.g., from 2 pg/m$^3$ for thulium and lutetium in samples S1 and S9 to 18 µg/m$^3$ for silicon in sample S5). The large variability in elemental composition is attributed to the inherently high variability associated with PM sources in greater Houston [13, 14, 55-58].

![Figure 1. Box plot of elemental concentrations in PM$_{10}$ over the study duration. The box encompasses the 25% and 75% percentiles, the whiskers span 1.5 times the interquartile range, and outliers are shown as diamonds. Inside each box, the horizontal line is the median value and the hollow square symbol (□) is the average value.](image-url)
Crustal elements (e.g., Na, Mg, Al, Si, Ca, Ti, and Fe) dominated in aerosols, suggesting that aeolian resuspension was their important source. Importantly, rare earth elements (REEs) were consistently detected, which exhibited abundances following the Oddo–Harkins rule with even atomic numbered REEs being more abundant than their immediate neighbors with odd atomic numbers except for samarium anomalies (i.e., La < Ce > Pr < Nd > Sm > Eu < Gd > Tb < Dy > Ho < Er > Tm < Yb > Lu). This trend validated the importance of crustal material as a dominant source of aerosols during the study period. Numerous main group (e.g., Li, Be, Ga, As, Sn, Sb, and Pb) and transition metals (e.g., V, Cr, Co, Ni, Cu, Zn, Mo, and Cd) were detected, many of which are known air toxics.

**Source apportionment**

Chemical Mass Balance (CMB) modeling estimated North African dust as the dominant source during the sampling campaign contributing 29 ± 22 percent (7–57 percent) over the study duration. Other local sources quantified included resuspended local soil/road dust as the second most dominant source, contributing 31 ± 16 percent (13–63 percent) over the study duration. Both these are predominantly derived from crustal materials, validating the observations made in the previous section regarding aeolian resuspension of aerosols. Construction activities and vehicular emissions also had significant impacts averaging 9 ± 6 percent (3–24 percent) and 11 ± 5 percent (3–20 percent), respectively. Although oil combustion modified vanadium atmospheric chemistry as described in the previous section, it contributed only negligibly to PM$_{10}$ mass 0.18 ± 0.25 percent (0–0.8 percent). Sea salt was another minor contributor to mineral matter, contributing an average of 2.3 ± 3.8 percent (0–9.7 percent). The relevant source contribution estimates from CMB modeling are summarized in Figure 2.

![Figure 2. Chemical mass balance modeling results for daily samples (S1–S9) depicting relevant source contribution estimates. Vehicular emissions are shown in red hatches and construction activities are depicted in blue hatches.](image-url)

**Bacterial community structure**

Following quality control, 16S rDNA gene sequencing generated an average of 30,981 reads per sample, varying between 2,005 and 65,154 reads. There were 848 bacterial OTUs recorded over the sampling campaign, representing 17 phyla, 46 classes, 117 families, and 121 genera.
Figure 3. (Top panel) The relative abundances of bacterial phyla over the nine-day sampling campaign. (Bottom Panel) The relative abundances of the top 30 bacterial genera, including the portion of reads unable to be classified to the genus level (Unclassified) and other, over the nine-day sampling campaign.

**Fungal community structure**

ITS rDNA gene sequencing resulted in a mean of > 109,000 counts per sample, which was significantly higher and less variable than that of bacteria. A total of 1345 fungal OTUs were identified, representing 4 phyla, 20 classes, 69 orders, 164 families, and 286 genera.
Potential human and environmental health impacts

The study of bioaerosols has become increasingly popular in recent years as scientists look not only at genetic but environmental risk factors for conditions like asthma, respiratory ailments, and certain cancers. Suspended bacterial and fungal cells that can maintain viability throughout short or long-distance transport can impact the surrounding climate, human health, plant health, animal health, atmospheric chemistry, and ecology.

Fungi responsible for invasive human diseases such as Aspergillus fumigatus, Fusarium spp. and Talaromyces spp. that are listed as being priority pathogens for global public health by WHO [16] appeared in nearly every sample along with all four most prominent allergenic fungal genera, viz. Alternaria, Cladosporium, Aspergillus, and Penicillium [59]. Their total relative abundance peaked in S3, the date of highest dust loading (Figure 5). Fungi form spores more readily than bacteria, resulting in their greater potential to remain viable even after exposure to wind and UV light when suspended. Because of their heightened durability, their potential impact to human health is emphasized.
Figure 5. Four prominent allergenic fungal genera and their relative abundances spanning the nine-day sampling campaign.

Numerous genera of plant or human pathogenic fungi including *Cercospora*, *Curvularia*, *Flavodon*, *Fomes*, *Fuscoporia*, *Funalia*, and *Magnaporthe* were also detected. Several pathogenic species peaked in abundance synchronous with North African dust including *Curvularia lunata*, *Rhizopus microsporus*, *Penicillium sclerotiorum*, *Phellinus gilvus*, *Exserohilum rostratum*, *Aureobasidium pullulans*, and *Curvularia trifolii*. We also detected saprobic fungi that play a critical role in carbon cycling such as *Basidiomycota*, *Neurospora*, *Peniophoraceae*, and the species *Phanerochaete chrysosporium* and *Trametes versicolor*.

Opportunistic human, plant, or animal pathogenic bacterial species were identified including *Escherichia coli*, *Propionibacterium acnes*, *Roseomonas mucosa*, and *Haemophilus parainfluenzae*. Several genera listed in WHO’s global priority pathogens list of multidrug and antibiotic-resistant bacteria and tuberculosis [15] were detected including *Achromobacter*, *Enterococcus*, *Haemophilus*, *Mycobacterium*, *Pseudomonas*, and *Staphylococcus*. Other pathogens measured included the Gram negative *Achromobacter*, *Caulobacter*, *Chitinophaga*, *Chryseobacterium*, *Clostridium*, *Corynebacterium*, *Gordonia*, *Listeria*, *Megasphaera*, *Nocardioides*, *Ochrobactrum*, *Propionivibrio*, *Sphingomonas*, *Stenotrophomonas*, and *Williamsia*, and the Gram positive *Actinomyces*, *Bacillus*, *Micrococcus*, *Nocardioidea*, *Propionibacterium*, *Roseomonas*, and *Rothia*. The plant pathogens *Erwinia*, *Rhizobium* (a nitrogen fixer), and *Streptomyces* as well as the filamentous cyanobacterium *Planktothrix* capable of seeding harmful algal blooms were also detected.

(Poly)extremophilic and thermophilic genera of bacteria (*Chroococcidiopsis*, *Deinococcus*, *Hydrogenophilus*, *Meiobacter*, and *Saccharomonospora*) and fungi [60] (*Alternaria*, *Aspergillus*, *Chaetomium*, *Cladosporium*, *Coprinopsis*, *Pencillium*, *Rhizopus*, *Scytalidium*, *Stemphylium*, *Talaromyces*, *Thanatephorus*, *Thermomyces*, *Thielavia*, and *Wallemia*) were also present.

An incomplete list of the potential pathogens noted in our dataset is included in Table 2 and Table 3 below.
### Table 2. Bacterial species present in the samples with potential pathogenicity.

<table>
<thead>
<tr>
<th>Species</th>
<th>Average Abundance</th>
<th>Impact</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Escherichia coli</em></td>
<td>0.76%</td>
<td>Animal</td>
<td>Avian pathogen</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Human</td>
<td>Human intestinal pathogen</td>
</tr>
<tr>
<td><em>Propionibacterium acnes</em></td>
<td>0.16%</td>
<td>Human</td>
<td>Opportunistic: acne vulgaris, endocarditis, endophthalmitis, prosthetic joint infections</td>
</tr>
<tr>
<td><em>Roseomonas mucosa</em></td>
<td>0.05%</td>
<td>Human</td>
<td>Opportunistic/nosocomial infector: peritonitis, bacteremia, endocarditis, endophthalmitis</td>
</tr>
<tr>
<td><em>Haemophilus parainfluenzae</em></td>
<td>0.02%</td>
<td>Human</td>
<td>Opportunistic: upper respiratory tract infections, urogenital infections, gastroenteritis, endocarditis</td>
</tr>
</tbody>
</table>

### Table 3. Fungal species identified in the dataset with potential pathogenicity.

<table>
<thead>
<tr>
<th>Species</th>
<th>Average Abundance</th>
<th>Impact</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Fomes fasciatus</em></td>
<td>1.38%</td>
<td>Plant</td>
<td>Wood decay</td>
</tr>
<tr>
<td><em>Funalia floccosa</em></td>
<td>1.14%</td>
<td>Plant</td>
<td>White rot</td>
</tr>
<tr>
<td><em>Curvularia lunata</em></td>
<td>1.03%</td>
<td>Plant</td>
<td>Tomato early blight, leaf spot</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Human</td>
<td>Phaeohyphomycosis</td>
</tr>
<tr>
<td><em>Tropicoporus tropicalis</em></td>
<td>0.49%</td>
<td>Plant</td>
<td>White rot</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Human</td>
<td>Keratitis, Mycosis</td>
</tr>
<tr>
<td><em>Schizophyllum commune</em></td>
<td>0.38%</td>
<td>Plant</td>
<td>Wood rot</td>
</tr>
<tr>
<td><em>Choanephora cucurbitarum</em></td>
<td>0.37%</td>
<td>Human</td>
<td>Lung infection</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Plant</td>
<td>Seedling rot, blight, fruit rot, leaf wilt, flower rot, stem necrosis, leaf spot</td>
</tr>
<tr>
<td><em>Rhizopus microsporus</em></td>
<td>0.33%</td>
<td>Human</td>
<td>Rhinocerebral mucormycosis, lung infection</td>
</tr>
<tr>
<td><em>Aspergillus fumigatus</em></td>
<td>0.29%</td>
<td>Human</td>
<td>Aspergilosis</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Animal</td>
<td>Aspergilosis</td>
</tr>
<tr>
<td><em>Penicillium sclerotiorum</em></td>
<td>0.26%</td>
<td>Plant</td>
<td>Leaf spot, post-harvest decay</td>
</tr>
<tr>
<td><em>Phellinus gilvus</em></td>
<td>0.17%</td>
<td>Plant</td>
<td>Wood decay</td>
</tr>
<tr>
<td><strong>Exserohilum rostratum</strong></td>
<td>0.15%</td>
<td>Plant</td>
<td>Leaf spot, rice brown spot, lettuce root rot</td>
</tr>
<tr>
<td>--------------------------</td>
<td>-------</td>
<td>-------</td>
<td>---------------------------------------------</td>
</tr>
<tr>
<td><strong>Stemphylium herbarum</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Brown spot, leaf spot</td>
</tr>
<tr>
<td><strong>Penicillium sumatraense</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Post-harvest decay</td>
</tr>
<tr>
<td><strong>Aureobasidium pullulans</strong></td>
<td>&lt;0.1%</td>
<td>Human</td>
<td>Opportunistic: subcutaneous phaeohyphomycosis, fungemia, peritonitis, pneumonia</td>
</tr>
<tr>
<td><strong>Fusarium oxysporum</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Stem rot, wilt, dieback</td>
</tr>
<tr>
<td><strong>Phanerochaete chrysosporium</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>White rot</td>
</tr>
<tr>
<td><strong>Amphobotrys ricini</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Gray mold</td>
</tr>
<tr>
<td><strong>Penicillium georgiense</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Post-harvest decay</td>
</tr>
<tr>
<td><strong>Curvularia trifolii</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Blight, leaf spot</td>
</tr>
<tr>
<td><strong>Cladosporium sphaerospermum</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Leaf spot</td>
</tr>
<tr>
<td><strong>Tilletia barclayana</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Rice/grain smut</td>
</tr>
<tr>
<td><strong>Hyphodermella rosae</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>White rot, dry fruit rot</td>
</tr>
<tr>
<td><strong>Nigrospora oryzae</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Leaf spot, leaf blight</td>
</tr>
<tr>
<td><strong>Aurantiporus fissilis</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Wood decay</td>
</tr>
<tr>
<td><strong>Purpureocillium lilacinum</strong></td>
<td>&lt;0.1%</td>
<td>Human</td>
<td>Opportunistic: hyalohyphomycosis, ocular infections</td>
</tr>
<tr>
<td><strong>Wallemia sebi</strong></td>
<td>&lt;0.1%</td>
<td>Human</td>
<td>Subcutaneous phaeohyphomycosis</td>
</tr>
<tr>
<td><strong>Trichothecium roseum</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Fruit rot, post-harvest decay, root rot, dieback, leaf spot</td>
</tr>
<tr>
<td><strong>Periconia macropinosa</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Leaf necrosis</td>
</tr>
<tr>
<td><strong>Aspergillus penicillioides</strong></td>
<td>&lt;0.1%</td>
<td>Human</td>
<td>Opportunistic: subcutaneous infections, keratomycosis, aspergillosis</td>
</tr>
<tr>
<td>-------------------------------</td>
<td>--------</td>
<td>-------</td>
<td>---------------------------------------------------------------</td>
</tr>
<tr>
<td><strong>Pilidium concavum</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Tan-brown rot, fruit rot, leaf necrosis</td>
</tr>
<tr>
<td><strong>Saccharomyces cerevisiae</strong></td>
<td>&lt;0.1%</td>
<td>Human</td>
<td>Opportunistic in immunocompromised patients: vaginitis, blood stream infections, essential organ infections</td>
</tr>
<tr>
<td><strong>Eutypa leptoplaca</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Grapevine pathologies; dieback</td>
</tr>
<tr>
<td><strong>Rhizopus arrhizus</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Root rot, soft rot, post-harvest rot</td>
</tr>
<tr>
<td><strong>Favolus gramocephalus</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>White rot</td>
</tr>
<tr>
<td><strong>Biatriospora mackinnonii</strong></td>
<td>&lt;0.1%</td>
<td>Human</td>
<td>Eumycetoma, cutaneous phaeohyphomycosis</td>
</tr>
<tr>
<td><strong>Gjaerumia minor</strong></td>
<td>&lt;0.1%</td>
<td>Human</td>
<td>Keratitis</td>
</tr>
<tr>
<td><strong>Antrodia pini-cubensis</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Brown rot</td>
</tr>
<tr>
<td><strong>Trametes versicolor</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>White rot</td>
</tr>
<tr>
<td><strong>Punctularia strigosazonata</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>White rot</td>
</tr>
<tr>
<td><strong>Tolypocladium inflatum</strong></td>
<td>&lt;0.1%</td>
<td>Animal</td>
<td>Insecticidal</td>
</tr>
<tr>
<td><strong>Monocillium indicum</strong></td>
<td>&lt;0.1%</td>
<td>Animal</td>
<td>Lymphadenitis, splenitis</td>
</tr>
<tr>
<td><strong>Quambalaria cyanscens</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Smut</td>
</tr>
<tr>
<td><strong>Cordyceps bassiana</strong></td>
<td>&lt;0.1%</td>
<td>Animal</td>
<td>Insecticidal</td>
</tr>
<tr>
<td><strong>Trichaptum abietinum</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Wood decay</td>
</tr>
<tr>
<td><strong>Thermomyces lanuginosus</strong></td>
<td>&lt;0.1%</td>
<td>Human</td>
<td>Opportunistic: endocarditis</td>
</tr>
<tr>
<td><strong>Malassezia restricta</strong></td>
<td>&lt;0.1%</td>
<td>Human</td>
<td>Opportunistic: endocarditis, pneumonia, dermatitis</td>
</tr>
<tr>
<td><strong>Bjerkandera adusta</strong></td>
<td>&lt;0.1%</td>
<td>Human</td>
<td>Bronchopulmonary mycosis</td>
</tr>
<tr>
<td><strong>Waitea circinata var. circinata</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Brown ring patch</td>
</tr>
<tr>
<td><strong>Phlebia chrysocreas</strong></td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Heartrot</td>
</tr>
<tr>
<td>Neopestalotiopsis foedans</td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Leaf spot</td>
</tr>
<tr>
<td>---------------------------</td>
<td>-------</td>
<td>-------</td>
<td>-----------</td>
</tr>
<tr>
<td>Phlebiopsis gigantea</td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Wood decay</td>
</tr>
<tr>
<td>Ceratobasidium ramicola</td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Dieback, leaf blight, leaf rot, damping off, root rot</td>
</tr>
<tr>
<td>Rigidoporus ulmarius</td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>White rot</td>
</tr>
<tr>
<td>Postia caesia</td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Brown rot</td>
</tr>
<tr>
<td>Exidia glandulosa</td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Wood decay</td>
</tr>
<tr>
<td>Botryosphaeria dothidea</td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Blueberry canker, brown rot, pear ring rot, kiwi soft rot</td>
</tr>
<tr>
<td>Gonatobotryum apiculatum</td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Leaf spot, blight</td>
</tr>
<tr>
<td>Fibroporia radiculosa</td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Wood decay, brown rot</td>
</tr>
<tr>
<td>Daldinia eschschoitzii</td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Wood decay</td>
</tr>
<tr>
<td>Gloeophyllum trabeum</td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Wood decay, brown rot</td>
</tr>
<tr>
<td>Thanatephorus cucumeris</td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Rice sheath blight, tomato seedling damping off, potato black scurf, stem canker, root rot</td>
</tr>
<tr>
<td>Phaeoacremonium croatiense</td>
<td>&lt;0.1%</td>
<td>Plant</td>
<td>Citrus tree diseases</td>
</tr>
</tbody>
</table>

**Conclusions and Recommendations**

Although we were able to quantify transportation through chemical analyses, microbiome composition was not as clear an indicator for particulate matter sourcing, potentially due to the ubiquity of certain microorganisms in the environment and limitations in genetic sequencing and identification. Our sampling campaign was incidentally overwhelmed by African dust and was fraught with multiple sources of particulate matter, making it difficult to exclusively assign microbial findings to the transportation sector.

Future studies should involve longer time-scales including when regional sources dominate aerosol composition in Houston. Furthermore, assessing viability (culturability) of microorganisms present in air samples is needed to better gauge the impact to ecosystem and human health.

**Outputs, Outcomes, and Impacts**

It is important to recognize that our sampling campaign unexpectedly coincided with a large African dust event. Although our primary objective was to determine emissions from the transportation sector, we also captured the role of long-range transported dust on PM\(_{10}\) levels along with its elemental and microbiological composition. Since this was a relatively “fundamental” research effort (i.e., not oriented towards technology development), our
findings are not directly amenable for technology transfer. Having said that, our sampling, elemental analysis, and microbial sequencing and interpretation tools can be applied to other studies of transportation aerosols. Some important take home messages were deduced, as itemized below.

**Research Outputs, Outcomes, and Impacts**

- A manuscript based on these data has been published after peer-review: Sourav Das, Alyvia McEwen, Joseph Prospero, Daniel Spalink, and Shankararaman Chellam (2023). *Respirable Metals, Bacteria, and Fungi during a Saharan-Sahelian Dust Event in Houston, Texas*. Environmental Science & Technology, https://doi.org/10.1021/acs.est.3c04158 [18].

**Technology Transfer Outputs, Outcomes, and Impacts**

- This is the first of its kind North American dataset simultaneously characterizing a wide suite of metals and microbial (bacterial and fungal) diversity in respirable PM.
- Because we captured Saharan dust, our work has findings that have broader implications for microbial ecology associated with long-distance dispersal, Earth’s radiation budget, climate, hydrology, and public health.
- Several metals and pathogenic microorganisms were present at elevated levels at a busy intersection, demonstrating increased human exposure to airborne toxics at this location. Cumulatively, direct vehicular emissions and resuspended soil/road dust contributed about 40 percent of the measured respirable PM mass. These findings can be extrapolated to other heavily trafficked intersections suggesting direct links between the portion of respirable PM arising from transportation and human health.
- Importantly, metagenomically identified bacteria and fungi do not imply that they are viable. Hence, future research needs to complement our preliminary findings by monitoring viability (culturability) to better assess microorganisms’ role in ecosystem and animal/human health.
- Lesson plans communicated to elementary school students inculcated interest in environmental issues related to the transportation sector.

**Education and Workforce Development Outputs, Outcomes, and Impacts**

Our undergraduate students coordinated multiple outreach activities regarding traffic-related air pollution to elementary and high school students throughout the project duration with mentorship from Dr. Chellam.

**Camp BUILD**

On the weekend of January 15, 2022, two of our undergraduate assistants, Alyvia McEwen and Elisabeth Gerstacker, helped lead an air pollution demonstration for Camp BUILD. Camp BUILD is a Zachry Department of Civil and Environmental Engineering initiative created to familiarize high school upperclassmen with basic engineering concepts, the Texas A&M University campus, and Civil and Environmental Engineering faculty. Camp BUILD places an emphasis on reaching first-generation college students and engineers from underserved school districts across Texas. Over the weekend, 20 high school students participated in campus tours, admissions presentations, and hands-on demonstrations covering each of the subdisciplines within civil engineering, including the demonstration prepared by our team.

For the experiment, a candle was lit, placed in front of an air hose, and extinguished. Students were positioned 1 meter downstream of the candle with a PM monitor, and they recorded PM$_{10}$ values using a handheld PM monitoring device at indicated time intervals for two minutes past when the candle was extinguished. The procedure was then repeated for a measurement of 2 meters downstream. Students divided out the necessary
responsibilities including equipment manager, timekeeper, and data collector. For the demonstration, students were divided into teams of five, which allowed everyone to take ownership of the experiment and practice multiple roles. After all groups had taken the measurements, the students were tasked with graphing the PM$_{10}$ levels over time for both configurations and comparing results.

Students seemed to enjoy seeing how seemingly invisible pollutants, a light smoke trail from the candle, evoked a dramatic response from the PM monitor, even at short times and large distances away from the source. They could see the numbers climbing on the PM monitor and hear the alarms progressing from a low to moderate hazard, to unhealthy for sensitive groups, then to severe hazard. It was stressed to the students that the burning candle represented larger-scale sources of incomplete combustion, including tailpipe emissions from vehicles. Students understood that residential areas close to busy highways or manufacturing facilities will experience different air quality issues than residential areas in more protected settings. This was presented as one of the reasons for erecting walls and planting trees in neighborhoods near highways (i.e., to physically exclude particulate matter). We discussed the differences between point and nonpoint sources of air pollution and noted that the students' graphs showed how air quality changes depending on distance from a point source and time since its emission. Overall, the outreach activity served to introduce environmental engineering as an important field of study to incoming first-generation college students, as well as provide teaching experience for our undergraduate assistants who are considering graduate studies.
Figure 6. (Top panel) Students take PM$_{10}$ measurements at directed time and distance intervals after extinguishing a candle. (Bottom panel) Students work in teams to translate data into a graph of PM$_{10}$ concentrations over time.

Southwood Valley Elementary School

On April 13, 2022, Alyvia McEwen and Sorya Meyer demonstrated air pollution experiments to the fourth-grade students at Southwood Valley Elementary School in College Station. Most students enrolled in this elementary school (61 percent) are underrepresented minorities and nearly the same percentage (55 percent) of the student body is classified as economically disadvantaged by the Texas Education Agency, ensuring that our STEM-related experiments reached a diverse set of students. We performed two 90-minute presentations in the cafeteria to all fourth graders who were present numbering approximately 100 students and 5 teachers.

The first demonstration was designed to show how motor vehicles contribute to air pollution through tailpipe emissions. In this experiment, several candles were placed among groups of students, each representing fossil fuel combustion in an internal combustion engine. A handheld PM monitor was used to first record the ambient concentrations within the school cafeteria before lighting the candles. For each PM reading, we let a few students take the monitor and read out the PM$_{2.5}$ and PM$_{10}$ measurements as well as the monitor’s built in health indicator.
(“good,” “moderate,” “unhealthy for sensitive groups,” “unhealthy,” “very unhealthy,” and “hazardous”). Students took note that in the absence of burning candles, the cafeteria’s indoor air was deemed “good.” Next, students were instructed to ignite each candle, which acted as a surrogate for (incomplete) fuel combustion in an engine and a source for aerosols. Other students volunteered to extinguish the candle, and students reacted to the burnt smell and the visible trail of soot emitting from the wick. Students used the PM monitor to see how the indoor air quality changed after lighting the candle at varying distances from the source. They personally documented increasing PM$_{2.5}$ and PM$_{10}$ concentration values soon after extinguishing the candles, which triggered the monitor’s built-in alarm when aerosol levels reached “hazardous” levels close to the source. Students were fascinated by how drastically the particulate matter concentrations rose in a short period of time after the candle was burned and extinguished. They were able to connect increasing aerosol levels from the burning candle to gasoline and diesel combustion in motor vehicles. Additionally, we explained to the students based on these measurements how vehicles are a major contributor to poor air quality especially in large cities with poor public transportation systems and consequently that are highly dependent on private vehicles like many in Texas.

The second demonstration covered the topic of particle resuspension by vehicles (i.e., non-tailpipe emissions). To illustrate this, a small patch of the floor was covered with flour, leaves, soil, and other debris. Then, students volunteered to help drive an RC car back and forth over the patch. The students were able to watch the flour and debris get kicked up into the air by the RC car tires. They were able to understand that microscopic particles can be similarly resuspended by less intense motions, such as walking or even rubbing their hands together, and they were also able to consider how many particles are resuspended by heavy traffic. In both demonstrations, it was emphasized to the students that our air quality is affected daily by industry and traffic, and it has health consequences for people that live or work in locations closest to the pollutant sources. At the end of our demonstrations, students were asked to share what they learned, and their responses showed excitement and appreciation that STEM knowledge (with engineering applications) could tackle transportation and environmental issues that affect real people every day.

![Figure 7.](image-url) **Figure 7.** (Left panel) Students measure PM levels near a lit candle. (Middle panel) Students recognize an increase in PM levels after the candle is extinguished. (Right panel) Volunteers drive the remote-controlled car over the flour patch as they visualize tires kicking up particles.

**Navarro Elementary School**

On October 17, 2022, Alyvia McEwen, Sorya Meyer, and Elisabeth Gerstacker outreached to roughly 75 students at Navarro Elementary School in Bryan, Texas. About 35 percent of the student body has limited English proficiency, and roughly 88 percent of the students are classified as economically disadvantaged. The same two
demonstrations from the Southwood Valley Elementary School outreach were performed for the three fourth-grade classes at Navarro, including one class of English as a Second Language students.

Figure 8. (Top panel) Undergraduate assistant Alyvia McEwen explains how to read the handheld PM monitor for the candle-burning demonstration. (Bottom panel) Volunteers drive the remote-controlled car over the flour patch as they visualize tires kicking up particles.

References


